MOLECULAR DIAGNOSTICS: EXPANDING ACCESS TO DRUG-RESISTANCE TUBERCULOSIS TESTING THROUGH CULTURE-FREE NGS

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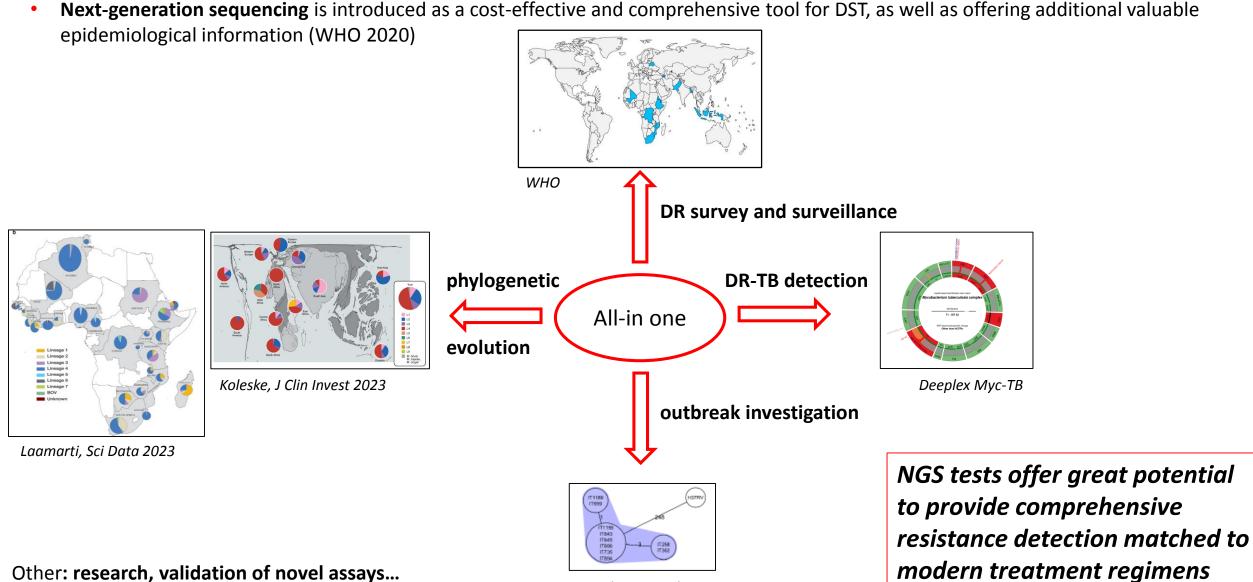
The need



- Only 2 in 5 people with MDR/RR-TB were known to have been diagnosed and enrolled on treatment in 2021
- Recently the WHO endorsed the use of a novel all-oral 6-month BPaLM regimen in people suffering from MDR/RR-TB, including those with
 additional resistance to fluoroquinolones (pre-XDR-TB) > better outcomes and quality of life, shorter duration
- However, there is need to expand access to drug-resistance testing, including to the drugs that constitute the best available regimens recommended for drug-susceptible and drug-resistant TB
- The current WHO-recommended rapid diagnostics (WRDs) cannot detect resistance to all the drugs in these two types of regimens in a single test to inform treatment decisions

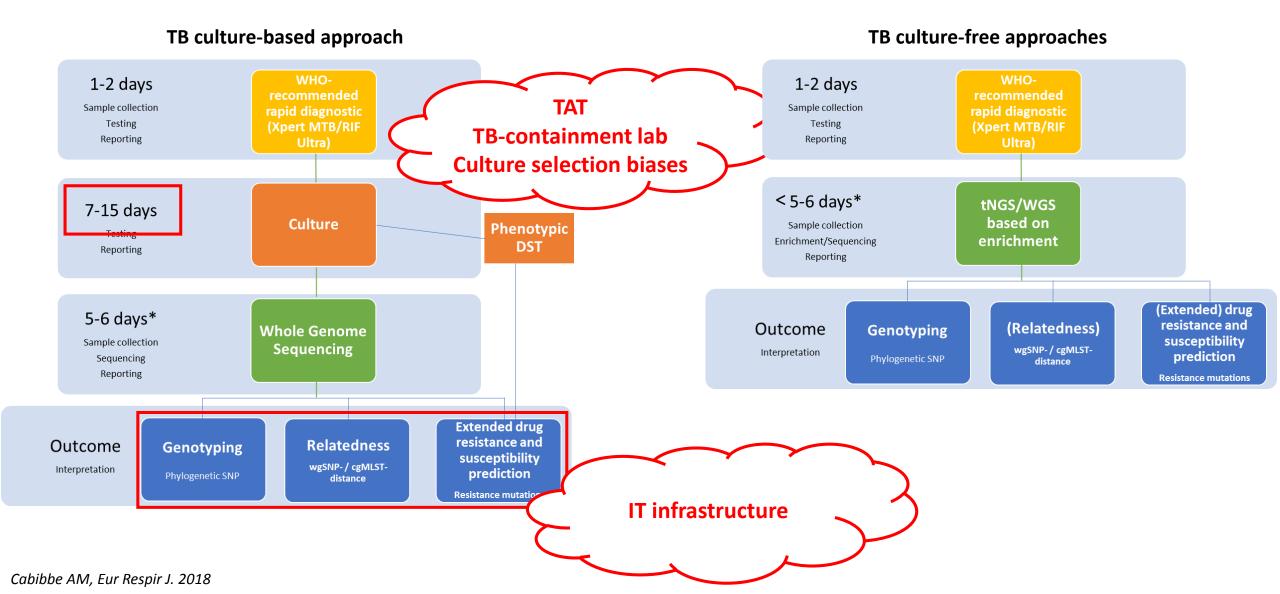
Increasing access to early and accurate diagnosis using a molecular WHO-recommended rapid diagnostic test

The solution (?)



Ridom SeqSphere

The NGS-based workflows



The challenge for TB culture-free NGS approaches

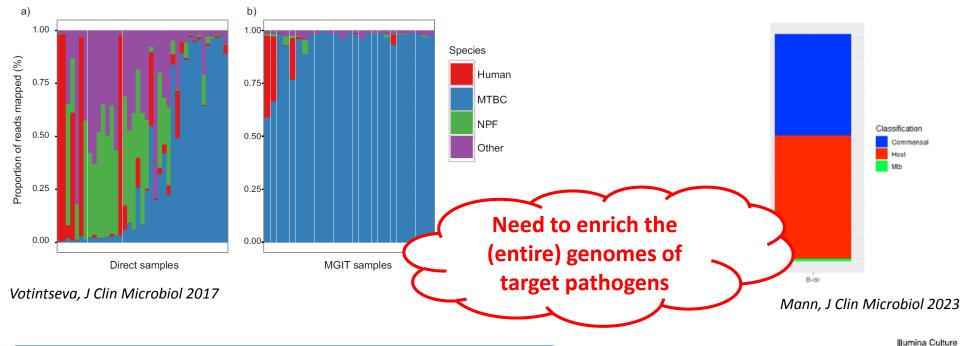
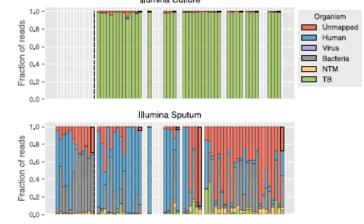


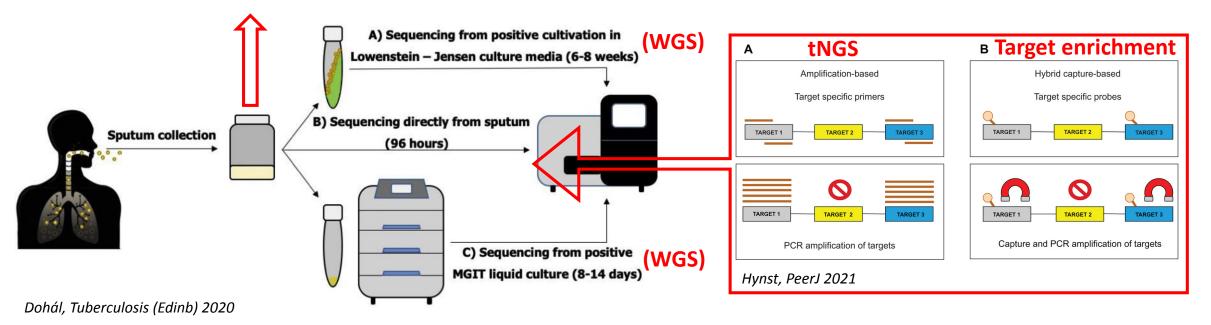
Table 1	Sample characteristics a	and sequencing results.
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Sample	ZN grade	DNA concentration in extract (µg/mL)	Total no. reads	% reads aligning to human genome
K1	3+	27.8	989,442	73.71
K2	3+	2.28	2,170,640	78.46
K3	2+	71	1,617,808	99.3
K4	2+	250	1,204,408	97.22
K5	2+	7.7	1,537,676	74.17
K6	2+	48.8	2,411,708	97.47
K7	1 +	25	2,818,238	50.59
K8	1+	0.63	1,851,892	20.29



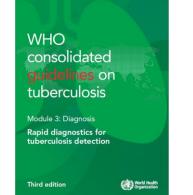
NGS enrichment approaches

Enrichment of MTB cells, e.g.: Antibody-based systems; Magnetic carriers with immobilized affinity ligands; capturing onto liquid or solid surface

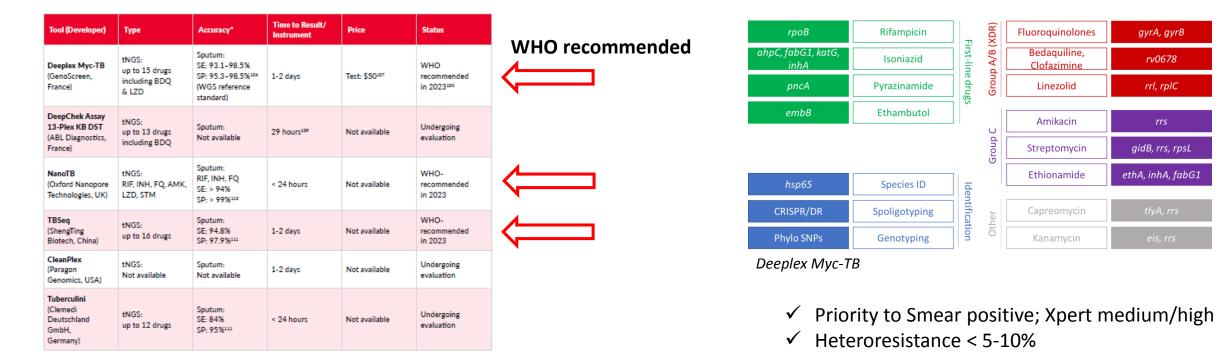


tNGS (1)

- Targeted next-generation sequencing was found to be accurate
- Targeted next-generation sequencing was found to be costeffective depending on context
- Targeted next-generation sequencing was found to be acceptable and implementable under routine conditions, despite inherent complexity



Among people with RR-TB: rifampicin, isoniazid, pyrazinamide, ethambutol, **fluoroquinolones, bedaquiline, linezolid**, clofazimine, amikacin and streptomycin



Pipeline Report 2023 (Treatment Action Group)

tNGS (2)

Strengths	Weaknesses	
✤ Multi-purpose, multi-disease	 Need of genotypic-phenotypic associations 	
✤ Suitable from various sample types	Turnaround time depends on sample referral and sequencing capacity/multiplexing	
 Rapid (faster turnaround times than conventional pDST testing) 	Start-up costs	
 Kit-based and user-friendly analysis tools (improved standardization) 	 Currently not feasible at peripheral level 	
Deep level of genetic information enabling "precision"	Procurement and supply chain	
	Need of specialized and trained personnel	

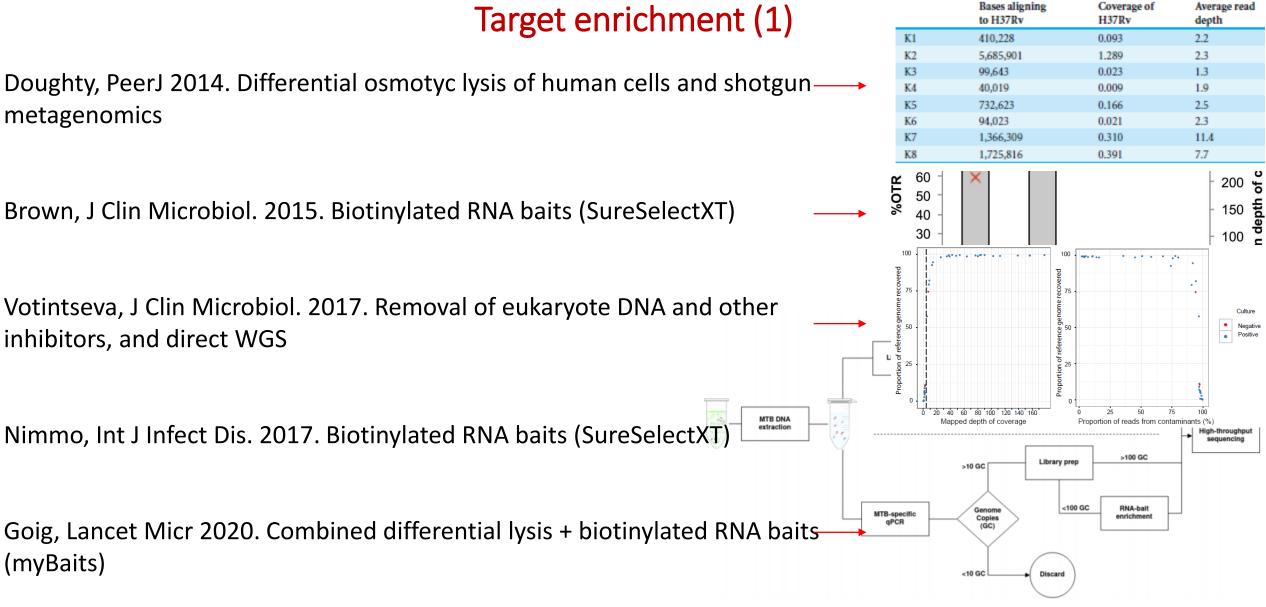
Opportunities	Threats	
Less phenotyping in routine testing	 Borderline mutations 	
 High predictive value for drug- resistance 	 Confidence-grading of mutations requires large and representative datasets 	
 Huge research on innovative NGS technologies 	 Support to clinicians 	
Development of lists of confidence- graded mutations reflecting on routine Nucleic Acid Amplification Tests	Not all resistance mechanisms can be explored (e.g., gene expression, structural changes)	
Interrogates resistance to additional anti-microbials not routinely tested in national algorithms	 Information technology (IT) infrastructure 	
✤ Research outcomes	 Cost-effectiveness to be demonstrated 	
	 Efficient and timely results reporting 	
	✤ To achieve sustainability	

This table provides a Strengths, Weaknesses, Opportunities, and Threats (SWOT) analysis of the use of genomics in TB control and care. The SWOT analysis helps to identify strategies to maximize the advantages and mitigate the disadvantages of having tNGS capacity for TB implemented.

Target enrichment (1)

Under relaxed mapping conditions

Sample



Soundararajan, Tube 2020. Mtb-specific tiling oligonucleotide probes

Target enrichment (2)

Wash

DNase

Benzonase

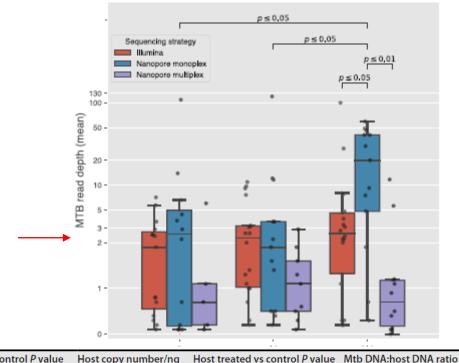
George, J Clin Microbiol. 2020. DNA thermo-protection buffer during heat inactivation and direct WGS.

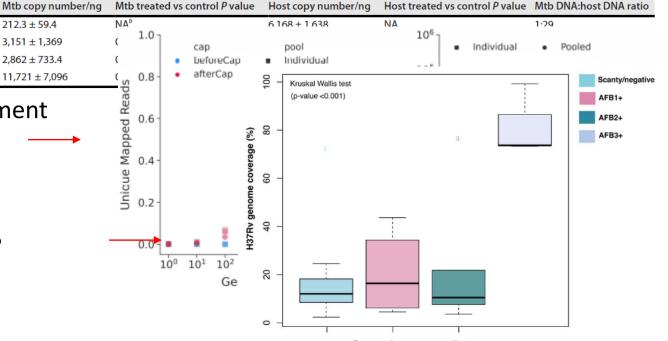
Nilgiriwala, J Clin Microbiol. 2023. DNA thermo-protection buffer and direct WGS on Illumina or ONT.

Mann, J Clin Microbiol. 2023. Benzonase/DNase and Twist target

Sundararaman, mBio 2023: Circular Nucleic acid Enrichment Reagent synthesis (DNA baits)

Vasanthaiah, preprint 2024. TB-Bead kit and direct WGS





Smear microscopy grading

Take-home messages

Critical steps for direct MTB WGS:

- Sputum / sediment sample input (ideally the whole volume)
- Selection / enrichment of bacterial cells (ideally MTB) over other population (e.g. human) present in the direct sample
- Selection / enrichment of bacterial DNA (ideally MTB) in the direct sample (recovery rate), quality of extracted MTB DNA (absence of inhibitors for PCR or enzymatic reactions of library prep protocols)
- Handling: number of steps / automation / costs

Current stage:

- ✓ tNGS is standardized (WHO approved commercial kits) but doesn't enable extended analyses
- ✓ Enrichment of whole genomes would enable higher resolution: however, lenghty and expensive procedures; genome coverage not yet good enough for clinical deployment